



A Mini Review of the Potentials of *Chlorella* Species Microalga for the Production of Biocrude by Hydrothermal Liquefaction (HTL) Operation

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ABSTRACT

The operation of hydrothermal liquefaction (HTL) of biomass is an enticing technique for the production of biocrude, which can serve as a viable alternative energy source to the growing universal energy demand and the pollution problem caused by fossil fuels. This article reviewed research publications that describe the elemental composition of Chlorella species biomass, biochemical analysis of the feedstock, the effects of the conditions of operating the Chlorella species biomass conversion, hydrothermal liquefaction mechanisms, separation technique, and the products of hydrothermal liquefaction operation. Nevertheless, the modification of the chemical and biochemical compositions of Chlorella specie biomass to yield higher-quality biocrude was not researched. Also, other biocrude separation techniques, such as mechanical separation, were not researched after the hydrothermal liquefaction of the Chlorella specie biomass. This information can help in making decisions on the route where research should be directed, towards in-depth study, improvement of yield of the biocrude more than other products, and quality of the derived biocrude by the operation of hydrothermal liquefaction of Chlorella species biomass.

Keywords: Biocrude; *Chlorella* specie; hydrothermal liquefaction; microalgae; separation technique

1.0 INTRODUCTION

About 88% of energy around the world that is utilized by humans is obtained from fossil fuels (Guo et al., 2015), but fossil fuels are non-renewable, produce greenhouse gases during combustion, and pollute the environment (Song et al., 2015). The utilization of microalgae globally for consumption and for microalgae products to be used as food supplements, dietary supplements, medicine, cosmetic industries, functional foods, and biofuels has greatly increased its production (Tang & Suter; Bito et al., 2020). The rapid urban development and industrialization in the past decades have resulted in the degradation of water quality, which has become a threat to public health (Gollakota et al., 2018), hence the need for alternative sources of fuel that are renewable (Tareket et al., 2022). *Chlorella* species are among the most studied microalgae, and they are widely cultivated for biotechnological applications (Aigner et al., 2020). Because of the properties of rapid growth rate, high oil content, high productivity of biomass, and renewability associated with *Chlorella* species, they are being considered as possible feedstock for the production of biocrude in comparison to other terrestrial biomass (Eboibi, 2019). The genus *Chlorella* is a single-cell microalgae, and the matured cells of *Chlorella* species are spherical and sometimes oval (Aigner et al., 2020), which contain a huge number of various functional compounds such as 12–18% lipid (Phusuntiet al., 2017), 50–60% protein, and 15–20% carbohydrate, zinc, growth hormones, calcium, potassium, vitamin E, B₁, B₂, C, B₆, magnesium, folic acid, free biotin, and chlorophyll (Tianet al., 2022). *Chlorella* microalgae can be found virtually in all geographic locations (Aignere et al., 2020). Presently, *Chlorella* species are made up of three varieties, which are *Chlorella lobophora*, *Chlorella vulgaris*, and *Chlorella sorokiniana* (Bito et al., 2020). *Chlorella sorokiniana* is a smaller species that was first isolated in 1953 by Sorokin but was originally thought to be a thermo-tolerant *Chlorella pyrenoidosa*, which is strange. Over 100 strains of *Chlorella* species have been described, with more than 20 characterized (Lizzulet et al., 2018). *Chlorella pyrenoidosa*, referred to as the subject of many scientific research studies, is now called *Chlorella sorokiniana* (Bito et al., 2018). *Chlorella* strains are reported to possess the ability to adapt to different environmental conditions and several carbon forms because they can be cultivated heterotrophically, autotrophically, mixotrophically, and photoheterotrophically (Piasecka & Baier, 2022).

Hence, renewable energy, where biomass is used as a source of energy, has attracted widespread attention (Hao et al., 2021). The B₁₂ content of *Chlorella* species products reported in the literature varies greatly from <0.1 to 400 g/100 g dry weight. Among all *chlorella* species grown in an open culture, the B₁₂ content is much higher in *Chlorella pyrenoidosa* than in *Chlorella vulgaris*. The B₁₂ is not essential for *Chlorella* species growth (Bito et al., 2020). Biomass refers to organic substances like animals, plants, and microorganisms, as well as materials derived from the metabolism and excretion of these organisms, with the exception of fossil fuels and their derivation (Rabacalet et al., 2017). Microalgae biomass has a higher calorific value and a lower density and viscosity than biomass obtained from plants. It accounts for the world's fourth-most consumed energy source after crude oil, natural gas, and coal (Cao et al., 2017). The organic composition of the biomass of *Chlorella* specie can be converted, extracted, and processed into fuels by chemical and physical techniques (Hao et al., 2021). This conversion can be carried out by hydrothermal operation, but there are several techniques that are used for the conversion of microalgae biomass into liquid fuels; these can be either biochemical or thermochemical techniques (Kumar et al., 2018). The types of these techniques are presented in Figure 1. Thermochemical methods include hydrothermal carbonization (HTC), hydrothermal gasification (HTG), and hydrothermal liquefaction (HTL), depending on the ranges of pressure and temperature at which the reaction is carried out (Duo et al., 2019). Hydrothermal operation involves the biomass undergoing different thermochemical pathways at different temperature and pressure conditions to form solid fuel, liquid fuel, and gaseous fuel in hot compressed water (Hao et al., 2021).

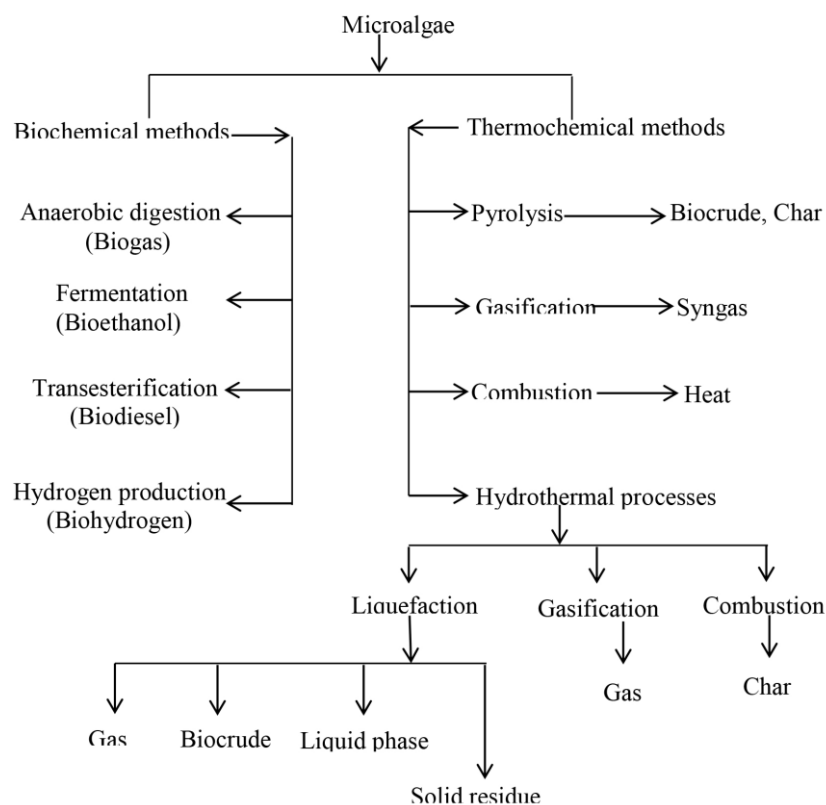


Figure 1 Flow chart of the biochemical and thermochemical methods

HTL produces biocrude, gases, solid residue phase, and aqueous phase in the presence of compatible catalyst and solvent at about 200–400 °C temperature (Xue *et al.*, 2016; Hao *et al.*, 2021), 0–28 MPa pressure (Hu *et al.*, 2019), and 5–60 minutes of residence time (Eboibi, 2019). The process of converting biomass to bio-crude involves the hydrolysis operation, degradation process, decarboxylation process, deamination, repolymerization, and other possible reactions (Hao *et al.*, 2021). The desired liquid product of HTL, which is biocrude, has high carbon and energy values that are similar to those of conventional petroleum (Eboibi, 2019), but biocrude has a high heteroatom content, a large acid value, a low higher heating value (HHV), and a high viscosity (Hao *et al.*, 2021).

Zhang *et al.* (2013) worked on biocrude production from the biomass of *Chlorella pyrenoidosa* by hydrothermal liquefaction using ethanol and water as solvents. The operation was carried out at 280 °C for about 60 minutes and 0.69 MPa. They reported a bio-crude yield of about 41.7% with an HHV of 31.1 MJ/kg when water was used as a solvent, but when ethanol was used alone as a solvent, the bio-crude yield increased to 51.1% with an HHV of 31.7 MJ/kg. Hague *et al.* (2022) reported the atmospheric effect and modified plasma on the HTL of *Chlorella* using DCM as a solvent. The HTL operation was carried out at about 240–250 °C with a residence time of 0–15 minutes using SiO₂/Al₂O₃ as a catalyst. They carried out the HTL operation at different conditions, giving an average total yield of biocrude of 49.83%. He *et al.* (2018) worked on the production of biocrude from the biomass of *Chlorella* by hydrothermal liquefaction before upgrading it catalytically. They used a 415-ml reactor, 53.208g of *Chlorella* microalgae, 212.8 ml of deionized water, DCM as a solvent, and Co Ni Mo W/γ-Al₂O₃ as a catalyst. The reactor was operated at a temperature of about 340 °C, a pressure of about 22 MPa, and a residence time of about 140 minutes. The total bio-crude yield was obtained as 28.2 wt%, and its higher heating value (HHV) was obtained as 31.9 MJ/kg. Jin *et al.* (2017) carried out the utilization of optimum biochemical components in *Chlorella* specie KR1 for biocrude production by hydrothermal liquefaction. The researchers used 95 ml of distilled water and 0.5g of lyophilized chlorella species KRI with DCM as solvent in a 42 ml high-pressure reaction vessel operated at 150 °C under autogenous pressure condition

They obtained a maximum biocrude yield of about and a higher heating value (HHV) of 27.62–33.16 MJ/kg. Nwanya *et al.* (2021) worked on the potential of the biofuel they produced from *Chlorella vulgaris*. They used 200g of dried *Chlorella vulgaris* and 300 ml of n-hexane in a Soxhlet extraction unit. The extraction was carried out for about six hours before recovering the oil in a rotary evaporator. They obtained a 37% yield of the biofuel, 45.125 MJ/kg as the calorific value, -7.667 °C as the pour point, and 56.67 °C as the cetane number. Chakraborty *et al.* (2013) carried out the isolation of α-glucan, a co-product of the derived biocrude, through the hydrothermal liquefaction operation of the *Chlorellasorokiniana* species with ethanol or water as solvents to extract lipid and polysaccharide from the microalgae. They performed sequential hydrothermal liquefaction at an optimum temperature of about 160 °C for the ethanol-insoluble polysaccharide and 200 °C for the water solvent, with a retention time of 10 g of alginate and the solvent (1:9). Rinanti and Purwadi (2019) researched the increasing yield of carbohydrates and lipids in *Chlorella vulgaris* (CH_{0.0052}N_{0.0013}O_{0.01472}P_{0.0589}) biomass, used for the production of biofuel. In their work, they used various concentrations of CO₂ ranging from 0.03% to 50% in the culture, and they obtained a carbon content of 0.57 g carbon/g biomass of *Chlorella* specie AG10002. They observed that the internal cellular energy storage of oil instead of carbohydrates actually slows down the reproduction rate of the microalgae. They used a photobioreactor for the microalgae cultivation, and the harvesting was carried out using the bioflocculation technique at room temperature, using microalgae as flocculants. The Fourier transform infrared (FTIR) patterns of the biomass of *Chlorella sorokiniana* were recorded to define the variety of functional groups involved in the microalgae. The waveband at approximately 3276.33 cm⁻¹ corresponds to O-H and N-H stretching vibrations in *Chlorella sorokiniana* biomass. The peaks obtained at 1636.14 cm⁻¹ and 1537.94 cm⁻¹ were attributed to amino group N-H stretching, while the C-OH stretching was assigned the band at 1031.52 cm⁻¹ (Embaby *et al.*, 2022). All these researchers used solvent for the extraction of the biocrude after the hydrothermal liquefaction operation, but none of these researchers used the mechanical method (gravity separation technique) for the recovery of the biocrude without the use of

solvent. Also, none of them studied how the yield and quality of the biocrude can be enhanced by modifying the composition of the biomass. Among these researchers, Zhang *et al.* (2013) obtained the highest biocrude yield at lower operating temperatures and pressures.

2.0 Elemental compositions of *Chlorella* species biomass

Generally, biomass consists of elements such as carbon (C), hydrogen (H), nitrogen (N), sulfur (S), and oxygen. The higher heating value of biomass can then be obtained with different models using the elemental compositions. These elements are explained below, and their percentage weight present in different *Chlorella* species is presented in Table 1 (Gollakota *et al.*, 2018; Gita *et al.*, 2019). The elemental composition of *Chlorella* species presented in Table 1 shows that there are similarities concerning carbon content (always above 40 wt%), hydrogen content (always above 5 wt%), and oxygen content (always above 20 wt%), with the exception of the work done by Rizzo *et al.* (2013), but there is some variance for the nitrogen content, especially for *Chlorella sorokiniana*, carried out by Chenet *et al.* (2014). In general, the elemental composition of the *Chlorella* species biomass reported falls within the standard range for all microalgae (see Table 1).

2.1 Carbon content of *Chlorella* species biomass

This is the most significant constituent of the *Chlorella* species biomass. Carbon obtained from the CO₂ in the atmosphere is taken in by the plant and becomes part of its composition during photosynthesis (Gollakota *et al.*, 2018). Carbon is the major contributor to the heating value of *Chlorella* species biomass, and during combustion, the carbon is converted into CO₂, which is released and can be channeled back to the cultivation system, where it becomes part of the photosynthetic process. Majorly, the content of carbon in the different biomasses is estimated through the compositions of hemiscellulose, lignin, and cellulose. Biomass rich in lignin has a carbon content that is higher than 50 wt% (Gollakota *et al.*, 2018). Different sources of carbon and their concentrations influence the structural composition of microalgal cellular lipids. When the CO₂ concentration is high, it promotes the accumulation of saturated fatty acids, but at low concentrations of CO₂, unsaturated fatty acid accumulation is induced (Jaiswalet *et al.*, 2020). Jabeenet *et al.* (2019) carried out thermogravimetric analysis on *Chlorella* algae using a Netzsch STA 449 F3 instrument, and they obtained a carbon composition of 52.1 wt%. Chang *et al.* (2014) worked on the characterization of *Chlorella* biomass activated carbon and reported that the carbon present is about 50.5 wt%. Zhenget *et al.* (2019) reported that the carbon/nitrogen ratio of *Chlorella vulgaris* cultivated in manure-free piggery wastewater had a significant effect on cell viability. They added that cell viability was improved when glycerol in the carbon/nitrogen ratios of 5:1, 25:1, and 125:1 were added compared with the control carbon/nitrogen ratio of 17:20. Their results show that the cell viability of the *Chlorella* species is enhanced by balancing the carbon/nitrogen ratios. Some microalgae species have the ability to use organic carbon sources instead of CO₂. *Chlorella* species heterotrophic growth can synthesize up to 45 wt% more carbohydrates and about 280 wt% more lipids compared to *Chlorella* species autotrophic growth (Jaiswalet *et al.*, 2020).

Jazrawiet *al.* (2015) worked on a double-staged hydrothermal liquefaction operation of a good protein-containing *Chlorella* species microalga. They obtained the carbon content of the biomass as 53.5 wt%. Gao *et al.* (2021) worked on the cultivation of mixotrophic and anaerobic hydrolysis for the treatment of *Chlorella* species wastewater in an anaerobic membrane photobioreactor and membrane photobioreactor. They obtained carbon contents of 0.38 and 0.47 g/g dry biomass for the anaerobic membrane photobioreactor and membrane photobioreactor, respectively. All these researchers at times determined the composition of carbon in the different *Chlorella* species biomass and also talked about the importance of carbon in the biomass and in the derived biocrude, but they did not research on how the carbon composition can be modified genetically for a higher heating value of the *Chlorella* species biomass and that of the derived biocrude.

2.2 Hydrogen content of *Chlorella* species biomass

This is another significant constituent of the *Chlorella* species biomass as an organic substance, as can be observed in carbohydrates, phenolic polymers, etc. During the combustion process of the *Chlorella* species biomass, the hydrogen component present is converted to water (H₂O), and this contributes greatly to the overall higher heating value (HHV) of the *Chlorella* species biomass. The content of hydrogen is normally lower in herbaceous biomass, that is, 5.5–6%, than in woody biomass, which is about 6–8% (Gollakota *et al.*, 2018). Microalgae biomass has been regarded as an important feedstock for hydrogen production because of its high growth rate and good protein and carbohydrate contents (El-Dalatony *et al.*, 2017). The use of hydrogen in the past decades has gained attention increasingly because of its high specific energy (142 MJ/kg) and its environmental friendliness. Jabeenet *et al.* (2019) carried out thermogravimetric analysis on *Chlorella* algae using a Netzsch STA 449 F3 instrument, and they obtained a hydrogen composition of 6.5 wt%. Gianget *et al.* (2019) reported that hydrogen production from *Chlorella* species biomass increased significantly from 1188 ml/l to 3055 ml/l when the biomass concentration was increased from 10 to 20 g/l. When the concentration of the biomass was further increased to 50 g/l, there was no significant improvement in hydrogen production, as it fluctuated within a narrow range of 2992 to 3180 ml/l. Jazrawiet *al.* (2015), who worked on a double hydrothermal liquefaction of a good protein-containing *Chlorella* species microalga, obtained the hydrogen content of the biomass as 7.4 wt%. The works of these researchers show that the significance of hydrogen composition in *Chlorella* species biomass cannot be overemphasized, both in the production of hydrogen and in increasing the HHV of *Chlorella* species biomass and the derived biocrude. Nevertheless, these researchers did not consider how the composition of hydrogen can be further enhanced genetically in the *Chlorella* species biomass.

2.3 Nitrogen content of *Chlorella* species biomass

Nitrogen constitutes a vital nutrient that is obtained from the *Chlorella* species biomass. The amount of nitrogen element present in the biomass contributes greatly to the process of degradation in fermentation or digestion, but during combustion of the biomass, nitrogen does not oxidize and therefore does not contribute to the overall heating values (Gollakota *et al.*, 2018). It is applied in compound form to the soil as fertilizer to enhance plant growth and its overall yield. The nitrogen element plays a significant role in the production of lipids and during cell growth. The total dry weight of the biomass of microalgae is composed of about 1–10% of nitrogen. The production of lipid in *Chlorella* species biomass increases under the condition of starvation of nitrogen element, which in turn reduces the total biomass of the *Chlorella* species, which reduces the overall yield of lipid (Ruet *et al.*, 2020). Jabeenet *et al.* (2019) carried out thermogravimetric analysis on *Chlorella* microalgae using a Netzsch STA 449 F3 instrument, and they obtained a nitrogen composition of 9.95 wt%. Jazrawiet *al.* (2015) worked on a double-staged hydrothermal liquefaction operation of a good protein-containing *Chlorella* species microalga. They obtained the nitrogen content of the biomass as 11.0 wt%. Gao *et al.* (2021) worked on mixotrophic cultivation and anaerobic hydrolysis for the treatment of *Chlorella* species wastewater in an anaerobic membrane photobioreactor. They obtained biomass nitrogen content of 0.140 and 0.092 g (dry biomass) for the anaerobic membrane photobioreactor and membrane photobioreactor, respectively. These different researchers, having reported that the nitrogen starvation condition increases the lipid production of microalgae while reducing the total biomass production, which will eventually affect the overall lipid yield, did not consider how the biomass production of *Chlorella* microalgae can be increased under the nitrogen starvation condition.

2.4 Sulfur content of *Chlorella* specie biomass

Sulfur is another significant nutrient similar to nitrogen in the structure of amino acids, enzymes, and proteins for improved plant growth. The sulfur content of the biomass of herbaceous crops is higher than that of woody biomass, thereby contributing to their high growth rate. The sulfur content of wood can be below the detection limit (0 wt%) and can be up to 1 wt% in exceptional cases, while it can be as high as 0.2 wt% or higher in herbaceous biomass but is most significant in gaseous emissions, corrosion, and syngas cleaning in gasification processes (Gollakota et al., 2018). Jabeenet et al. (2019) carried out thermogravimetric analysis on *Chlorella* species of microalgae using a Netzsch STA 449 F3 instrument, and they obtained a sulfur composition of 0.55 wt%. Jazrawiet al. (2015) worked on a double-staged hydrothermal liquefaction of a good protein containing *Chlorella* specie microalga. They obtained the sulfur content of the biomass as 0.5 wt%. Since it was reported by these researchers that the presence of sulfur in microalgae enhances the growth rate, they did not work on how the sulfur content of the *Chlorella* species can be further enhanced for a higher growth rate and possible ways that the oxide formation of sulfur can be reduced when it occurs. Ferreira & Soares-Dias (2020) studied the pyrolysis of *Chlorella vulgaris* over carbonate catalysts and obtained the sulfur content of the microalgae biomass as 0.7 wt%.

2.5 Oxygen content of *Chlorella* specie biomass

When considering the biomass chemical composition of *Chlorella* species, the oxygen element is very significant. The amount of oxygen present in the biomass influences the heating value. The oxygen content in phenolic compounds is difficult to break so as to enhance the heating values. The measurement of the oxygen content is not done directly; it is estimated by subtracting the concentrations of carbon (C), hydrogen (H), nitrogen (N), sulfur (S), and ash from 100, whether wet or dry biomass (Gollakota et al., 2018). Jazrawiet al. (2015) worked on a double-staged hydrothermal liquefaction operation of a *Chlorella* species microalga containing a good amount of protein. They obtained the oxygen content of the biomass as 27.6 wt%. Jabeenet et al. (2019) carried out thermogravimetric analysis on *Chlorella* algae using a Netzsch STA 449 F3 instrument, and they obtained an oxygen composition of 30.9 wt%. Khooet al. (2020) reported that the amount of oxygen decreases when the temperature of the hydrothermal liquefaction is elevated from 180 to 250 °C, with the raw microbial biomass having an oxygen content of 59.7 wt%. Ferreira & Soares-Dias (2020) researched the pyrolysis of *Chlorella vulgaris* over carbonate catalysts and obtained the oxygen content of the *Chlorella vulgaris* biomass as 35.1 wt%. Zhang et al. (2019) researched the property variation of microalgal biomass, its comparison, and its characterization, and they obtained the *Chlorella* specie biomass oxygen content as 31.4 wt%.

Table 1. Elemental composition of *Chlorella* species

Species	Carbon (wt%)	Hydrogen (wt%)	Nitrogen (wt%)	Surphur (wt%)	Oxygen (wt%)	HHV (MJ/kg)	Citation
Microalgae	35-57	6–9	2 - 12	~1.5	24-35	-	Gollakota et al., (2018)
Chlorella	52.31	6.99	10.14	0.86	29.70	-	Jin et al., (2017)
Chlorella KR1	53.62	8.10	2.59	0.61	35.08	24.61	Jin et al.,(2017)
C. Pyrenoidosa	51.4	6.6	11.1	-	30.9	-	Zhang et al., (2013)
C. Pyrenoidosa	51.0	6.6	11.0	-	31.4	-	Gai et al., (2015)
Chlorella	40.31	5.99	9.14	0.76	27.74	17.31	Xu et al., (2019)
C. Pyrenoidosa	50.99	7.83	9.48	1.08	30.62	-	Peng et al., (2016)
C. Sorokiniana	50.42	7.89	2.91	0.23	37.44	21.63	Mao et al., (2012)
C. Vulgaris	47.7	7.5	8.4	0.50	35.9	22.03	Guo et al., (2019)
C. Vulgaris	48.5	7.0	8.5	0.2	35.0	20.2	Yang et al., 2018)
Chlorella sp.	46.1	6.1	6.7	0.4	19.1	-	Babich et al., (2011)
Chlorella sp.	50.2	7.25	9.30	nr	33.2	nr	Babich et al., (2011)
C. vulgaris	52.6	7.1	8.2	0.5	32.2	nr	Billier & Ross, (2011)
C. vulgaris	45.8	7.9	7.5	nr	38.7	nr	Chakinala et al., (2009)
C. vulgaris	42.51	6.77	6.64	nr	27.95	nr	Wang et al., (2013)
C. vulgaris	48.3	7.3	3.0	nr	32.9	nr	Figueira et al., (2015)
C. vulgaris	50.39	6.01	14.77	6.05	22.78	nr	Raheem et al., (2015)
C. vulgaris	50.0	7.1	5.8	0.53	33.8	nr	Peng et al., (2017)]
C. vulgaris	44.8	6.8	7.0	1.0	40.4	nr	Lopez-Gonzalez et al., (2014)
C. vulgaris ESP-31	53.01	8.67	3.26	nr	35.05	nr	Bach et al., (2017)
C. pyrenoidosa	51.2	6.8	11.3	0.7	30.7	nr	Gai et al., (2015)
C. sorokiniana CY1	40.32	7.38	2.61	nr	44.50	nr	Chen et al., (2014)
Chlorella	51.34	7.29	9.35	nr	31.45	23.09	Tarek et al., (2022)

3.0 Biochemical analysis of the *Chlorella* species biomass

The biochemical analysis of the *Chlorella* species biomass includes the determination of carbohydrate content, lipid content, moisture content, ash content, protein content, and other valuable bioactive compounds such as carotenoids (Safafaret al., 2016). These components are presented in Table 2. Zhang et al. (2014) determined the biochemical composition of *Chlorella pyrenoidosa* by Fourier transform infrared (FTIR) spectroscopy. They washed the cell pellets with deionized water twice and then suspended the cell pellets in fresh deionized water again at a concentration of approximately mg/mL (dry weight). A total of 200 µl of suspension was dropped on a KRS-5 window (30 x 5 mm) and dried at 40 °C in the vacuum oven. The transmittance of the spectra was determined between 400 and 4000 cm⁻¹ at a resolution of 4 cm⁻¹ with 32 scans on the equipment. Rani & Ojha (2021), in their research on *Chlorella sorokiniana*, obtained biomass yields of about 126 and 158 mg(l.d.) in synthetic wastewater and 319 and 548 mg(l.d.) in real tertiary wastewater under 12:12 h and 24:0 h light and dark regimes, respectively. Sakarika and Kornaros (2017) researched the lipid accumulation and kinetics of the growth of the cells of *chlorella vulgaris* and obtained a biomass yield of about 2.69 g/l. Huang et al. (2016), in their research work, achieved a stable biomass productivity of 103 g/m² after cultivation of *Chlorella vulgaris* in an attached system, which is about 30.4% higher than that cultivated when the system is suspended. Gao et al. (2021) worked on mixotrophic cultivation and anaerobic hydrolysis for the treatment of *Chlorella* specie wastewater in an anaerobic membrane and membrane photobioreactor. They obtained biomass production rates of 32.55 and 91.10 for the anaerobic membrane and membrane photobioreactor, respectively.

3.1 Lipid content of *Chlorella* species

Lipids constitute 5 to 40% of the *Chlorella* species biomass, which is mainly glycolipids, phospholipids, waxes, hydrocarbons, and fatty acids, which, when produced in the chloroplasts, are directed to the cell walls and cell membranes of the several organelles, as well as the chloroplasts themselves and the mitochondria. When *Chlorella* specie is cultivated in an unfavorable environment, the production of lipid may

be up to about 58% of the total composition of the *Chlorella microalgae* (Mobinet et al., 2019). Lipids form the main component that transforms into biocrude after the hydrothermal liquefaction operation of microalgae. Figure 2 shows the possible lipid chemical reaction pathways in the hydrothermal liquefaction operation. Lipids may be hydrolyzed by a non-catalytic method to quickly produce higher fatty acids and glycerols under hydrothermal liquefaction operations. Some of these higher fatty acids transform into hydrocarbons with long chains through the process of decarboxylation, which then transform into alkanes through hydrogenation. Hao et al. (2021) Some of the higher fatty acids transform into alcohols through the process of deoxygenation; the alcohols are further esterified with fatty acids to produce esters of fatty acids. Also, some of the higher fatty acids may react with the ammonia that is produced in the conversion process of amino acids to produce fatty acid amides. All these components make up the derived biocrude from biomass hydrothermal liquefaction operations (Kumar et al., 2016). The lipids of microalgae usually take the form of triacylglycerols (TAGs), which mostly have an aliphatic character (Barreiro et al., 2013). They are mostly non-polar, and they are made up of a glycerol backbone that is bonded to three fatty acids (Barreiro et al., 2013; Gollakota et al., 2018). Some of the major compositions of *Chlorella* species lipids are palmitic, oleic, and linolenic acids. The composition and content of the lipids of microalgae vary with different species, temperature, geographical location, season, salinity, light intensity, or a combination of these factors. Mobinet et al. (2019). *Chlorella vulgaris* and *Chlorella emersonii* can have more than 50% weight lipid content (Rani et al., 2018). At room temperature, the fats are generally not soluble in solvents, and as temperature changes, they tend to be polar. When TAGs are hydrolyzed, glycerol is one of the products obtained, and this glycerol can be processed to produce acetaldehyde, methanol, propionaldehyde, acrolein, ethanol, allyl alcohol, and formaldehyde, as well as a gas mixture (CO, CO₂ and H₂), when subjected to hydrothermal liquefaction (Gollakota et al., 2018). Within the profiling of lipids that has been carried out on *Chlorella* species, a higher concentration of fatty acids with 16 and 18 carbon atoms has been found (Table 2), and the specific heat capacity of some of the fatty acids is presented in Table 3.

It can be observed from Table 4 that most of the fatty acids present have carbon atoms and are not short-chain. These *Chlorella* species fatty acids with carbon atoms are favorable for health when consumed. It has been identified that 70.18% by weight of the *Chlorella* species fatty acids corresponds to saturated fatty acids (SFA), 16.85% by weight corresponds to monounsaturated fatty acids (MUFA), and about 8.72% by weight corresponds to polyunsaturated fatty acids (PUFA), while account of the balance was not reported, therefore fatty acids give a wide range in their lipid profile [69]. Zhang et al. (2014) researched the effects of carbon from organic sources on the biochemical composition and growth of *Chlorella pyrenoidosa*. In their report, the addition of 3.0 g/l galactose and glucose greatly reduced the content of lipid by 27.9 and 27.5%, respectively, while the addition of 5.0 g/l and 0.5–1.0 g/l galactose and glucose and 0.5–5.0 g/l fructose, starch, and disaccharide did not affect the lipid content significantly. They further reported that the effect glucose has on the lipid content of *Chlorella* species might be dependent on the specific strain and also dependent on the concentration of glucose.

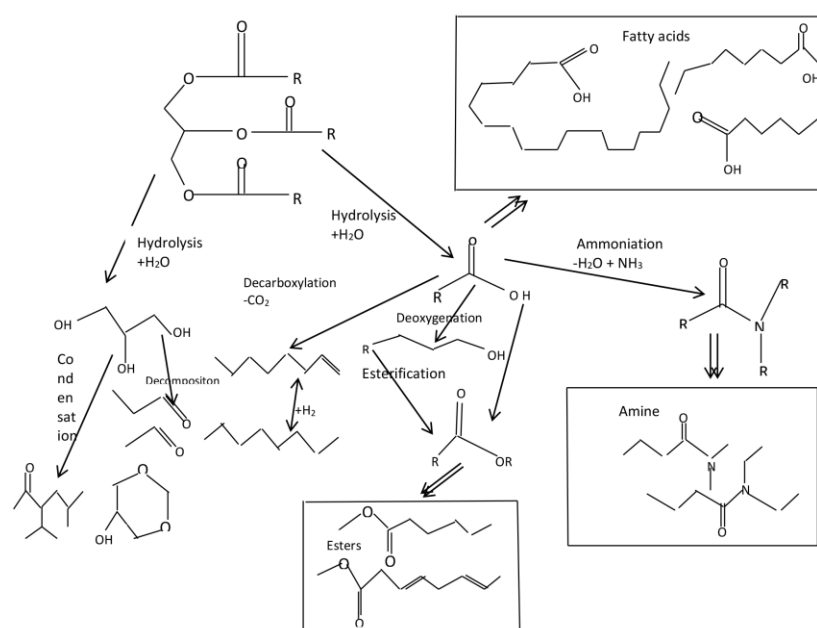


Figure 2: Lipids chemical reaction pathways in HTL

Table 2 Biochemical composition of *Chlorella* species (Safafar et al., 2016)

Species	Starting concentration	Nitrogen (mmol/l)	Biomass Conc. (g/l)	Lipid content (%dry weight)
<i>Chlorella vulgaris</i>	33.6 mM KNO ₃	33.6	11	15
<i>Chlorella vulgaris</i>	1.5 g/l NaNO ₃	17.6	nr	5.9
<i>Chlorella</i> sp.	0.025 g/l urea	0.8	0.464	66.1
<i>Chlorella vulgaris</i>	5 mM KNO ₃	5.0	0.4	15.9
<i>Chlorella vulgaris</i>	0.0003% KNO ₃	0.03	0.017	57.9
<i>Chlorella vulgaris</i>	10 mg/l nitrate	0.2	0.72	20
<i>Chlorella vulgaris</i>	70.02 mg/l nitrogen	5.0	0.86	30
<i>Chlorella vulgaris</i>	40 mg/l nitrate	0.6	0.48	60

Table 3 Specific heat capacity of triacylglycerols (Zhu et al., 2018)

Triacylglycerols	Specific heat capacity kJ/kg
Tristearin	1.938
Triolein	1.886
Trilinolein	1.813
Trilinolenin	1.765

Table 4 Fatty acids composition in *Chlorella* species (Coronado-Reyes et al., 2022)

Fatty acids	Carbon number	Percentage weight
Palmitic acid methyl ester	C _{16:0}	17.5
Palmitoleic acid methyl ester	C _{16:1}	3.27
Stearic acid methyl ester	C _{18:0}	15.39
Oleic acid methyl ester	C _{18:1}	13.87
Linoleic acid methyl ester	C _{18:2n6}	9.18
Lignoceric acid methyl ester	C _{24:0}	22.11

3.2 Protein content of *Chlorella* species

This is one of the significant constituents of microalgae biomass, and it is made up of many chains of peptides that grade into polymers of amino acids (Gollakota et al., 2018). High protein content is one of the nutritional characteristics of these microalgae species, i.e., about 60%, which also includes most of the essential amino acids. This amount of protein in *Chlorella* species is three times more than what is available in beef, which is considered to be one of the most concentrated sources of protein (Rani et al., 2018). The yield of amino acids when subjected to HTL is low because they undergo decarboxylation, which produces carbonic acids and amines; they also undergo deamination to form organic acids and ammonia (Hao et al., 2021). These products from the reactions may then be repolymerized to form aromatic ring structures such as nitrogen heterocyclics (pyrrole or indole), phenols, and long-chain hydrocarbons. Proteins are the major source of nitrogen in the derived biocrude (Barreiro et al., 2013). Since the protein of microalgae has a low molecular weight, once the cells of the *Chlorella* species have been disrupted, the protein can be easily digested, but peptides extracted from *Chlorella* species have significant protection against cell damage (Rani et al., 2018). The total protein composition of *Chlorella vulgaris* is about 42-48% dry weight in the biomass produced. Of this percentage, about 20% adhere to the cell wall and serve as structural and transport roles for the cell. About 50% of the proteins are intracellular proteins, which function mainly as enzymes, while the remaining 30% develop the microalgae after being secreted into the extracellular medium. The composition of the amino acids of the *Chlorella* species that has been identified contains isoleucine, histidine, leucine, lysine, threonine, methionine, valine, and phenylalanine (Coronado-Reyes et al., 2022). The chemical reaction pathways involve the breakdown of the amide bonds (Figure 3) in the peptide chain before it combines with water molecules, which will generate amino acids (Hao et al., 2021). Some of the amino acids formed (-COOH) decarboxylate to carbon dioxide (CO₂) and amine compounds. Part of the amine compounds produced tends to form water-soluble compounds containing nitrogen, and they are partitioned in the aqueous phase of the hydrothermal liquefaction. These processes improve the quality of the biocrude produced effectively by removing the oxygen and nitrogen contents present in it. Furthermore, some of the compounds of amine are also converted into compounds that are soluble in water, such as pyrazine and pyrrole in biocrude (Zhang et al., 2017). The total protein content present in the *Chlorella* species microalgae can be estimated by the standard Lowry method. In this method, bovine serum albumin (BSA) can be used as a standard. A spectrophotometer is then used to measure the absorbance of the sample using a maximum wavelength of 750nm. The protein concentration present in the *Chlorella* species biomass can be calculated by using the equation below:

$$\text{Total protein \% (w/w)} = \frac{CVD}{X} \times 100 \quad (1)$$

Furthermore, some of the compounds of amine are also converted into compounds that are soluble in water, such as pyrazine and pyrrole in biocrude (Zhang et al., 2017).

The total protein content present in the *Chlorella* species microalgae can be estimated by the standard Lowry method. I

Where *C* represents the concentration of protein in the *Chlorella* species biomass, *V* represents the volume of lysis buffer added to the *Chlorella* species biomass that has been freeze-dried for suspension, *d* represents the dilution factor, and *X* represents the amount of biomass (mg) (Rani & Ojha, 2021). El-Sheekh et al. (2020) worked on the *Chlorella vulgaris* and obtained the estimated protein content as 39.85 wt% of dry mass, which lies in the range of 33–46% when the *Chlorella vulgaris* was cultured under different light regimes. They also obtained the protein content as 23.7 wt% by mass when cultured in Bold basal medium (BBM), but under nitrogen limitation conditions. In this method, bovine serum albumin (BSA) can be used as a standard. A spectrophotometer is then used to measure the absorbance of the sample using a maximum wavelength of 750nm. The protein concentration present in the *Chlorella* species biomass can be calculated by using the equation below:

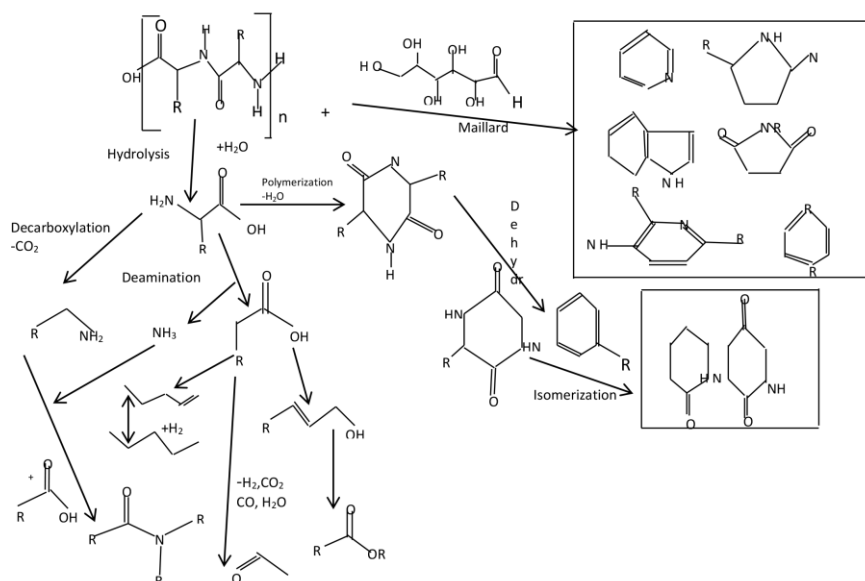


Figure 3: Protein chemical reaction pathways in hydrothermal liquefaction

3.3 Carbohydrate content of *Chlorella* specie

Chlorella specie contains carbohydrates in different forms, such as starch, cellulose, and some reducing sugars. The starch is found in the chloroplasts of the *Chlorella* species, consisting mainly of amylose and amylopectin. Cellulose is part of the cell wall of *Chlorella* species whose glycosidic bond is β 1-3; consequently, this can be used as input for the formation of functional products that contribute to human health (Coronada-Reyes *et al.*, 2022). Carbohydrates are mostly decomposed into materials that are polar and water-soluble, rather than non-polar hydrocarbons that eventually dissolve in the aqueous phase of the hydrothermal liquefaction process of the biomass (see figure 4). Consequently, a small amount of carbohydrates converts into biocrude during the process (Hao *et al.*, 2021). The total content of carbohydrate present in the *Chlorella* microalgae biomass can be evaluated by the method of phenol-sulfuric acid. In this method, glucose can be used as a standard for phenol-sulfuric acid. This is followed by the use of a spectrophotometer to measure the absorbance of the sample using a maximum wavelength of 490nm. The carbohydrate concentration present in the *Chlorella* species biomass can be calculated by using the equation below:

$$\text{Total (\% carbohydrate content)} = \left(\frac{X}{0.1} \right) \times 100 \text{ mg of glucose} \quad (2)$$

X (mg of glucose) is the absorbance at 0.1 ml of the sample test (Rani & Ojha, 2021).

The characterization and identification of the sugars present in *Chlorella* species is best carried out with high-performance liquid chromatography (HPLC), and with this equipment, the composition of the *Chlorella* species cell walls has been identified to result from a mixture of rhamnose, xylose, glucose, arabinose, galactose, and mannose, where rhamnose is the most abundant (Coronada-Reyes *et al.*, 2022).

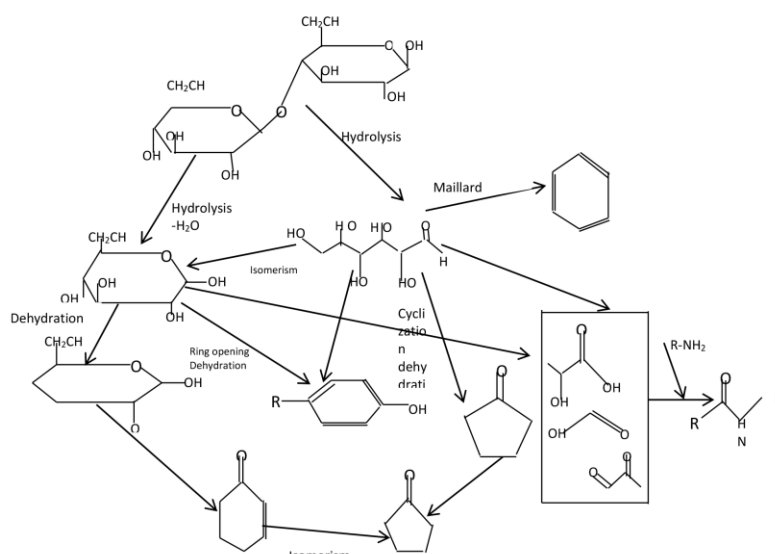


Figure 4: Carbohydrates chemical reactions pathways in hydrothermal liquefaction

Table 5: Biochemical analysis of *Chlorella species*

Species	Ash (wt%)	Moisture (wt%)	Proteins (wt%)	Lipids (wt%)	Carbohydrate (wt%)	Citation
Chlorella	6.5	5.7	60	2.8	25	He et al., (2018)
Chlorella KR1	5.13	-	22.17	-	36.12	Jin et al., (2017)
Chlorella Pyrenoidosa	5.6	-	71.3	-	22.0	Zhang et al., (2013)
Chlorella Pyrenoidosa	5.6	6.3	71	-	22	Gai et al., (2015)
Chlorella	-	-	60.1	2.9	20.2	Xu et al., (2019)
Chlorella Pyrenoidosa	5.81	-	70.65	-	21.75	Prng et al., (2016)
Chlorella Sorokiniana	-	-	16.8	24.7	-	Mao et al., (2012)
Chlorella Vulgaris	6.8	3.6	51.9	23.6	9.2	Guo et al., (2019)
Chlorella Vulgaris	3.0	-	54.0	18.7	24.3	Yang et al., (2018)
Chlorella sorokiniana	nr	nr	40.50	22–24	26.8	Hossain et al., (2019)
Chlorella sp. FC2ITG	nr	nr	22–41	15–54	18–46	Sajjadi et al., (2018)
Chlorella vulgaris	nr	nr	51–58	14–22	12–17	Rinanta & Purwadi, (2019)
C. sorokiniana	5.4	8.5	51.7	22.7	15.5	Niccolai et al., (2019)
C. vulgaris Allma	9.3	4.9	56.8	16.9	5.9	Niccolai et al., (2019)
Chlorella vulgaris	nr	nr	nr	9.23	5.7	Rinana & Purwadi, (2019)
Chlorella vulgaris	9.50	5.83	51.45	12.18	11.86	Coronada-Reyes et al., (2022)

4.0 Effects of operating conditions on the HTL process of *Chlorella specie* biomass

Commercialization and industrialization of biocrude production by hydrothermal liquefaction require a reduction in the production of gaseous phase, aqueous phase, and solid residue to obtain high yields and high-quality biocrude. Consequently, because of the difference in reaction parameters, there is considerable variation in the percentage yield of the biocrude obtained from biomass and the quality of the biocrude (Xue et al., 2016). Therefore, the effects of biomass/water ration, holding time, effect of biomass feedstock, effects of reaction temperature on the operation, heating rate, effect of operating pressure, effect of catalyst on the biomass, and effect of atmosphere (Tian et al., 2014)

4.1 Temperature effect on the HTL operation

One of the deciding parameters in the hydrothermal liquefaction operation of chlorella species microalgae is temperature (Xue et al., 2016). At subcritical temperature conditions, an increase in the reaction temperature will increase the yield of biocrude by fracturing the chemical bonds of the biomass (Kumar et al., 2016). A typical subcritical temperature at which hydrothermal liquefaction is performed is in the range of 200–400 °C. Xue et al., (2016). The role of temperature in hydrothermal liquefaction is defined by the competitive reaction between hydrolysis and depolymerization. At temperatures below 220 °C, the dominant reaction is hydrolysis, but the highest yield of biocrude is normally observed in the 250–375 °C temperature range, while at temperatures >375 °C, gasification becomes active, which promotes gaseous product formation (Hu et al., 2019). Duan et al. (2013) presented in their report that the maximum yield of biocrude was obtained from *Chlorella pyrenoidosa* at 350 °C; this corroborates the report by He et al. (2018) for biocrude production from *Chlorella species* at 340 °C. Zhang et al. (2013) obtained the maximum yield of biocrude at 280 °C for *Chlorella pyrenoidosa*, which is close to the report by Hague et al. (2022), which reported the optimum yield of biocrude from *Chlorella specie* to have occurred in the temperature range of 240–250 – 250°C. Therefore, the reaction temperature of biocrude production by hydrothermal liquefaction operations of microalgae varies largely among feedstock species.

4.2 Effect of residence time on HTL

This can be defined as the duration at which the hydrothermal liquefaction operation will be maintained at a designated temperature, outside the heating and cooling times. Higher temperatures require a shorter residence time. Generally, the residence time for the hydrothermal liquefaction operation is (Barreiro et al., 2013). Several studies have been carried out on the residence time effect on the hydrothermal liquefaction of *Chlorella species* of microalgae for the production of biocrude. Duan et al. (2013) worked on the residence time effect on the conversion of *Chlorella pyrenoidosa* to biocrude at 350 °C and obtained maximum yield after 70 minutes. Zhang et al. (2013) obtained the maximum yield of biocrude from *Chlorellapyrenoidosa* after 60 minutes at 280 °C. Miao et al. (2012) reported the effect of reaction conditions on biocrude production and polysaccharide production from *Chlorella sorokiniana* by the operation of hydrothermal liquefaction. They obtained the maximum yield of biocrude at 240 °C after 20 minutes.

4.3 Effect of heating rate on HTL

This is the ratio of the operating temperature to the time taken for the operation. The effect of the rate of heating the microalgae is not mentioned in the literature, as only a few studies have been done on it; therefore, its effect on the yield of biocrude is still unclear (Barreiro et al., 2013). Studies on *Chlorella vulgaris* by Biller et al. (2011) show that increasing the rate of heating from 10 to 25 °C/min slightly decreases the percentage yield of biocrude. Zhang et al. (2013) reported that when the rate of heating increases from 5 to 140 °C/min, the yield of biocrude increases by 30% for the conversion of grassland perennials to biocrude by HTL. Consequently, there is a need for further studies on the heating rate of microalgae biomass conversion to biocrude by hydrothermal liquefaction.

4.4 Effect of *Chlorella* biomass/water ratio on HTL

The presence of water in the operation of hydrothermal liquefaction of *Chlorella species* biomass promotes pyrolysis and dehydration, which actually enhance the breakdown of intermediates and subsequently repolymerize the small molecules (Xue et al., 2016). Water also helps in stabilizing free radicals while improving the quality of the biocrude. In most investigations carried out, the ratio of biomass to water used varies. Eboibi (2018) used about 1:5 for biocrude production by hydrothermal liquefaction of *Tetraselmis specie*; Hague et al. (2022) used 1:5 for biocrude production from *Chlorella* by HTL; Kumar et al. (2018) used 1:6, 1:7, 1:8, 1:9, and 1:10 for the hydrothermal liquefaction of microalgae and obtained maximum biocrude yield at a ratio of 1:9. However, excess water concentration decreases biocrude yield because, when the water concentration is in excess, it easily splits the macromolecules of the algae into small gas molecules.

4.5 Effect of Catalyst on HTL of *Chlorella* species

Catalysts are significant substances in chemical reactions. Undoubtedly, they are very important in the rate of conversion of *Chlorella* specie biomass to bio-crude and in the composition and quality of the bio-crude obtained by hydrothermal liquefaction (Tianet *et al.*, 2014). In hydrothermal liquefaction (HTL) operations, the catalysts used can either be heterogeneous or homogenous catalysts (Guo *et al.*, 2015). Heterogeneous catalysts such as Pd, Pt, or Ru that are supported on C, Co Mo, Ni, Pt, Ni/SiO₂, supported on Al₂O₃, and zeolite present many setbacks like sintering, dissolution, poisoning, and intraparticle diffusion limitations (Barreiro *et al.*, 2013). Homogenous catalysts in the form of acids such as phosphoric, acetic, sulfuric, hydrochloric, and perchloric acids are the most commonly used acid catalysts for biomass hydrothermal liquefaction. Weak acids like formic and acetic acids are usually preferred as solvents rather than catalysts in HTL, but they lead to the production of biocrude with a high content of oxygen (Xue *et al.*, 2016), while the corrosive nature of strong acids hinders their applications in the industry. Homogenous catalysts in the form of alkali salts like KOH and Na₂CO₃, have a positive effect on microalgae

Table 6 Catalyst used for the production of biocrude by HTL

Catalyst used	microalgae species	microalgae HHV (MJ/kg)	Biocrude yield (wt%)	Reference
Na ₂ CO ₃	Botryococcus branunii	34.2	64	Ross et al., (2010)
Na ₂ CO ₃	Chlorella vulgaris	24.6	20	Biller et al., (2011)
Pt/Al ₂ O ₃	Chlorella vulgaris	22.2	34	Eboibi, (2018)
Na ₂ CO ₃	Chlorella vulgaris	22.2	35	Biller & Ross, (2011)
Pd/C	Chlorella sorokiniana	21.7	30	Mao et al., (2012)
—	Chlorella pyrenoidosa	21.3	39	Yu et al., (2011)

4.6 Effect of pressure on HTL of *Chlorella* species

Another significant factor that affects the production yield of biocrude from the biomass of *Chlorella* species is pressure. Pressure helps to keep water in a single phase during hydrothermal liquefaction under supercritical or subcritical conditions, thereby avoiding the enthalpy required for phase change in water (Xue *et al.*, 2016). The acid-catalyzed effect is increased as the water density increases, which in turn increases the release of more hydrogen ions (H⁺) from compressed water at high temperatures (Guo *et al.*, 2015). A common way of doing this is by using a vacuum pump to evacuate the air from the reactor and substituting it with nitrogen, making it oxygen-free (Tianet *et al.*, 2014).

4.7 Effects of the physical state and load of *Chlorella* microalgae

The initial state of *Chlorella species microalgae* at the point of loading affects the yield of biocrude production by hydrothermal liquefaction. The microalgae physical state can be in dried pulverized form (Nwanya *et al.*, 2021; He *et al.*, 2018), dried freeze pulverized cells mixed with water (Biller *et al.*, 2011), or the state of the microalgae cells as received after harvest (Mao *et al.*, 2012; Gai *et al.*, 2015). When the pulverized sample is used, the rate of extracting constituents may be changed, therefore affecting the total yield of the biocrude and its composition, but the reported data of the HTL experiment of microalgae as received from harvest is more meaningful for the total yield and quality of the biocrude (Barreiro *et al.*, 2013).

5.0 Hydrothermal liquefaction mechanism

The operation of hydrothermal liquefaction is carried out at a subcritical temperature of about 200–400 °C and a pressure of about in the presence of water to produce biocrude, biogas, aqueous phase, and solid residue as represented in Fig. 5, where bio-crude is the desired product and others as by-products (Hao *et al.*, 2021). During the operation, the biomass decomposes and depolymerizes into monomers or smaller units in the subcritical water through a catalytic or non-catalytic path to produce small molecules that rearrange through cyclization, condensation, and polymerization to produce biocrude (Mathimani & Mallick, 2019). The hydrothermal liquefaction pathway is made up of three major steps: the de-polymerization process, which occurs first, followed by the decomposition process, and the recombination process (Gollakota *et al.*, 2018). The selectivity of these hydrothermal liquefaction mechanisms can differ depending on the pH of the medium, the severity of the HTL operating conditions (temperature, pressure, ramping, and retention time), the type of solvent used for the extraction and its concentration, and the type and nature of the catalyst used (Basare *et al.*, 2021). Hague *et al.* (2022) researched the plasma modification effect on the hydrothermal liquefaction of *Chlorella* species using a catalyst. The *Chlorella* species they used was purchased from Stakich incorporated, while the catalyst (zeolite) with a surface area of 780 m²/g, was purchased from Alfa Aesar, and they used ethanol and dichloromethane as solvents. They carried out the hydrothermal liquefaction operation in a batch reactor fitted with a stirrer and an electric heater. They introduced 4.5g of the *Chlorella* species into the reactor, followed by the addition of 22.5 ml of deionized water and 17.75 ml of ethanol, respectively. 0.225g of the catalyst was added, and the sample tube was corked for the hydrothermal liquefaction (HTL) operation. After reaching 240 or 250 °C, it was allowed to remain for about 15 min. They carried out the extraction of biocrude from the final product with the addition of 25 ml of dichloromethane (DCM) to extract organic components from the liquid and solid products. They kept the two phases, which are immiscible (organic solvent-soluble and water-soluble), in a separating funnel overnight. The DCM was evaporated at 45 °C for 20 min, and the ethanol was evaporated at 78 °C for 40 min. They obtained a total biocrude yield at different conditions with an average of about 50.0 wt% with a dark color and high viscosity. A major drawback of the HTL is the high operating temperature and pressure, which result in high capital costs for the operation. The authors reported that biocrude can be produced economically by adding alcohol to the HTL water to reduce the high temperature and pressure and modifying a suitable heterogeneous catalyst. Nevertheless, the researchers did not report whether any other solvent can be used as a substitute for the high efficiency of biocrude production at a low cost.

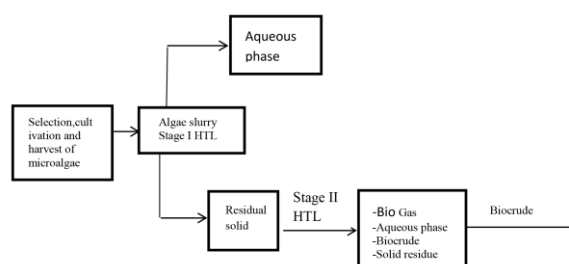


Figure 5: Diagrammatic representation of the entire process

5.1 Biomass depolymerization

Depolymerization is the first stage in which the bonds of the biocrude are broken by the substitution of molecules based on their chemical and physical properties, and the long-chain hydrocarbons produce the short-chain hydrocarbons (Mathimani & Mallick, 2019). Depolymerization solves the problems and inexorable properties of biomass from lignocellulose that imitate the natural process of fossil fuel formation (Gollakota et al., 2019). In these depolymerization or hydrolysis reactions, lipids are decomposed into glycerol and fatty acids; the proteins present are broken down to produce amino acids, while carbohydrates are broken down to produce monosaccharides (Huang et al., 2016). At this stage of hydrothermal liquefaction, no appreciable formation of biocrude takes place. If the factors affecting the HTL operation are adjusted in such a way that only the depolymerization step is affected, the effluent obtained can be further used in the fermentation process to produce fuels such as methane, hydrogen, and ethanol (Basaret et al., 2021). Depending on the characteristics of the substrate, the temperature range required for depolymerization at this level of HTL varies between 150 °C and 250 °C (Tianet et al., 2014). The depolymerization operation in the hydrothermal process is a widely researched technology in the pretreatment section of the hydrothermal field (Fan et al., 2016). This technology is primarily used to enhance the lignocellulosic hydrolysis of biomass or increase the biodegradability of municipal sludge (Cantero et al., 2015). Most substrates used in hydrothermal liquefaction, such as herbaceous plants and woody plants, cyanobacteria, microalgae, macroalgae, and manure, all contain various ratios of lignocellulose (Basaret et al., 2021). Depolymerization of the biomass of lignocellulosic produces different pentoses and hexoses together with monolignols. The monosaccharides produced from the depolymerization of *Chlorella species* biomass by hydrothermal means can contribute to the formation of biocrude by reacting with amino acids (Terrell et al., 2020).

5.2 Deposition of *Chlorella species* biomass monomers

This step of deposition of *Chlorella species* biomass monomers involves the loss of carbon dioxide molecules by decarboxylation, dehydration (which is the loss of water molecules), and withdrawal of the content of amino acids by deamination (Jena et al., 2015). The decarboxylation and dehydration processes aid in the oxygen withdrawal in the form of depolymerization of *Chlorella species* biomass and water, respectively, from the *Chlorella species* biomass, thereby reducing the oxygen content (Kumar et al., 2018). The *Chlorella species* biomass hydrogen bond is broken at the subcritical temperature and pressure of the water, forming polar monomers (Mathimani & Mallick, 2019). Smaller molecules of *Chlorella species* biomass that are hydrolyzed at high temperatures go through a series of thermal breakdowns such as decarboxylation, dehydration, decarbonylation, deamination, dehydrogenation, and some cleavages of the bonds. This decomposition reaction of various compound groups in HTL takes place in the 180–340 °C range (Hom-Diaz et al., 2015). In the operation of hydrothermal liquefaction, the first group of compounds to start decomposition is carbohydrates at about 180 °C. When the temperature increases above 200 °C, proteins and lipids begin to breakdown, and their decomposition will be total at about 300 and 640 °C, respectively (Chen et al., 2018). Total breakdown of lipids is not required in the hydrothermal liquefaction operation since some lipids, that is, fatty acids with long chains, are directly included in biocrude formation. The balance of the decomposition reactions is directly related to the efficiency of the HTL. The strength of the operating conditions of the hydrothermal liquefaction should be sufficiently high to aid the organic compounds in breaking down into biocrude-forming compounds. Also, it should not be too high to avoid decomposing biocrude into gases. In the application of the treatment of wastewater, decomposition reactions in the hydrothermal operation allow for the treatment of most pharmaceutical materials and the conversion of microplastics into biocrude (Dimitriadis & Bezeraiani, 2017). Carbohydrates, which are the first group to degrade in the hydrothermal operation, have different decomposition pathways that end with volatile basic alcohols, short-chain fatty acids, furan compounds, ketones, furanic alcohols, and furanic acids. With the exception of large furanic molecules, the breakdown of carbohydrates does not really contribute significantly to the formation of biocrude. However, the products from the decomposition of carbohydrates can form bigger molecules during recombination reactions. The second group that decomposes mostly in hydrothermal liquefaction operations is proteins. This decomposition pathway consists primarily of amino acid deamination and decarboxylation reactions (Basaret et al., 2021). The deamination reaction of amino acids is the primary source that produces ammonia in the aqueous phase of hydrothermal liquefaction. Consequently, protein-rich substrates in the hydrothermal liquefaction operation aqueous phase can have up to about 16 g/l ammonia with pH levels over 8 (Changiet al., 2015). The end products of the decomposition of proteins are mostly simple amines, amides, fatty acids with short chains, aldehydes, and N-heterocyclic compounds (Watson et al., 2020). N-heterocyclic molecules of protein decomposition are the only significant direct contributor to the formation of biocrude (Basaret et al., 2021).

5.3 Recombination of the reactive fragment of *Chlorella species* biomass

The recombination reaction is the last step in the hydrothermal liquefaction pathway. In this step, the fragments of the *Chlorella species* are repolymerized due to the absence of hydrogen compounds, and the large molecules formed in this recombination reaction contribute to the formation of biocrude at a temperature above 300 °C (Kumar et al., 2018; Basaret et al., 2021). Most of the organic molecular groups present in the biocrude, which are formed in the recombination process of long-chain fatty acids, are: amides, amines, aromatics, ketones, and esters. Also present in the biocrude are complex alcohol molecules, which come from the cyclization and hydration reactions of alkenes (Denielet et al., 2015). At this subcritical condition, the absence of hydrogen compounds is excessively large, and then the reactive fragments are repolymerized to produce char with a high molecular weight, which is referred to as the formation of coke (Gollakota et al., 2018). In the HTL operation, the Maillard type of reaction is one of the most significant recombination pathways for biocrude production. In this Maillard reaction, reducing sugars' carbonyl groups react with the free amino groups present in amino acids, which form nitrogenous polymers and melanoidins, which contribute to the production of biocrude (Qiu et al., 2019). Research carried out on model compounds revealed that if the Maillard reaction did not take place in a hydrothermal liquefaction operation, the biocrude yield from proteins and carbohydrates would be in the range of 7–18 wt%, while a biocrude yield of about 40 wt% was reported when the Maillard reaction occurred with the same carbohydrates and proteins in the hydrothermal liquefaction operation (Fan et al., 2018). Recombination of the small fragments also includes some of the biocrude compounds that form coke and eventually contribute to the formation of hydrochar (Basaret et al., 2021). But this coke formation can be reduced with the introduction of high-pressurized hydrogen gas during the recombination step (Barreiro et al., 2013). At this point, when the hydrothermal liquefaction conditions become severe, the biocrude begins to crack again, resulting in a decrease in process efficiency (Basaret et al., 2021).

5.4 Products of HTL from *Chlorella species* biomass

The end products of the hydrothermal liquefaction (HTL) of *Chlorella species* are the biogas, the aqueous phase, the biocrude fraction, and the solid residue. The total mass of these products is properly accounted for by carrying out material balance on them, together with the feedstock. These end products need to be separated in such a way that biocrude organic liquid, which is the desired product, will be obtained without much negative effect on its quality. The biocrude has an energy content of about 30 to 40 MJ/kg (Eboibi, 2019). The solvent extraction method is widely used for the separation operation, using a separating funnel. Dichloromethane (DCM), acetone, and chloroform are some

widely used organic solvents for the extraction of biocrude from the aqueous phase and solid residue (Shanmugamet et al., 2020). Hargue et al. (2022) found out that the yield of biocrude increased up to about 43.76% when treated with zeolite as a catalyst at about 240 °C, but the weight percentages of the gas phase, aqueous phase, and solid residue were not reported.

5.4.1 Gaseous product of the HTL of *Chlorella* specie biomass

A gaseous product is a co-product of hydrothermal liquefaction, which is usually obtained first, immediately after the system cools to room temperature. The gaseous fraction represents about 10% of the feedstock that was originally fed into the system (Hu et al., 2019). It is primarily made up of CO₂ and a small amount of C₂H₄, C₂H₆, CH₄, and H₂ (Tian et al., 2014). Depending on the hydrothermal liquefaction conditions, part of the nitrogen in the feedstock can be converted to ammonia, and both the ammonia and CO₂ produced can be recycled into algae cultivation medium (Tian et al., 2014; Wang et al., 2018). Gai et al. (2015) presented in their report that hydrothermal liquefaction of *Chlorellapyrenoidosa* at varying temperatures shows that the gas yield increased continuously as temperature increased but did not report the percentages of gas yield at the various temperatures. Zhang et al. (2013) reported that the gas phase of the HTL of *Chlorella pyrenoidosa* increases from to, respectively.

5.4.2 Aqueous phase of the HTL of *Chlorella* specie biomass

In the operation of all three types of hydrothermal processes, which are liquefaction, gasification, and carbonization, aqueous products are produced because they all involve the use of water (Tian et al., 2014). The composition of the aqueous phase produced from *Chlorella microalgae* by hydrothermal liquefaction includes PO₄³⁻, NH₄⁺, CH₃COO⁻ and some minerals such as K⁺, Na⁺ and Mg²⁺. Therefore, its recirculation plays a significant role in the *Chlorellamicroalgae* cultivation (Hu et al., 2019). Zhang et al. (2014) did not report the percentage yield of aqueous phase produced from *Chlorella pyrenoidosa* hydrothermal liquefaction operation, but Gai et al. (2015) presented that the percentage yield of the aqueous layer increases with increase in temperature, from 200 – 320 °C when *Chlorellapyrenoidosa* was subjected to hydrothermal liquefaction.

5.4.3. Biocrude phase of the HTL of *Chlorella* specie biomass

Biocrude is a dark viscous liquid similar to petroleum, but it is produced by hydrothermal liquefaction of biomass (Hu et al., 2019). The chemical and physical compositions of biocrude are largely influenced by the properties of the feedstock and the operating conditions of the system. Bio-crude is the desired product in the hydrothermal liquefaction operation, and the yield varies largely, even among the same strains and the same operating conditions (Barreiro et al., 2014). Biocrude is a complex mixture involving a large number of compounds and several distributions of molecular weight, and its analysis is commonly performed with the use of gas chromatographic equipment fitted with mass spectrometry (GC-MS) (Hu et al., 2019). Zhang et al. (2014) reported that the biocrude yield obtained by hydrothermal liquefaction of *Chlorella pyrenoidosa* using 50 wt% ethanol as solvent was 57.3 wt%, while Gai et al. (2015) also reported that the biocrude yield fell after an initial rise in the HTL of *Chlorella pyrenoidosa* from 200 to 320 °C. Xue et al. (2014) reported that when HZSM-5 was used as a catalyst, the biocrude yield of *Chlorella pyrenoidosa* was 34.02 wt%, and when Ce/HZSM-5 was used, it was 49.87 wt%, while Zhang et al. (2013) reported that the yield of biocrude from the hydrothermal liquefaction of *Chlorella pyrenoidosa* was 70.8 wt% at 240 °C and 47.1 wt% at 300 °C.

5.4.4 Solid residue of the HTL of *Chlorella* specie biomass

This is primarily made up of inorganics and trace organic matter remaining after the HTL operation (Tian et al., 2014). The solid residue percentage yield of HTL is largely determined by the content of ash in the feedstock, and it is usually observed to be less than 10 wt% because of the low ash content of *Chlorella microalgae*. HTL solid residue might contain products of protein that are degradable, which suggests it might be useful as a feed additive for animals because of its possible high nutritional value (Hu et al., 2019). Zhang et al. (2014) reported that the solid residue yield obtained by HTL of *Chlorella pyrenoidosa* using different concentrations of ethanol as a solvent produces solid residue ranging from 9.42 wt% to 17.4 wt%. Gai et al. (2015) reported that the percentage yield of solid residue decreases gradually as the temperature of the operation increases from 200 to 320 °C, while Zhang et al. (2013) reported that for non-catalytic HTL at about 200 °C and 220 °C, the solid residue obtained was 59.5 wt% and 37.5 wt%, respectively.

6.0 Challenges in the Production of Biocrude from *Chlorella* Species Biomass and Future Directions

The unique advantages of the production of biocrude from the *Chlorella species* feedstock and the nature of its environmental friendliness make the hydrothermal liquefaction operation of *Chlorella microalgae* a promising technology for biorefinery. The prospects and challenges for the HTL of *Chlorella microalgae* studies and production are as listed:

1. Inability to improve the amount of chemical and biochemical compositions such as carbon, hydrogen, which contributes to the HHV of the biomass, and lipids, which are the main components that transform into biocrude, while reducing the amount of the components, does not favor the production of biocrude nor enhance its quality. This can be achieved by genetic modification of the *Chlorella* species during cultivation.
2. Application of solvent extraction techniques for the separation of the biocrude, which require the use of chemicals and additional techniques for the separation of chemicals. Mechanical separation techniques, such as gravity separation, can be employed.

7.0 CONCLUSION

The hydrothermal liquefaction operation of *Chlorella* species appears to be a very attractive technology for the production of biocrude, but this technology is still in its early stages. Several studies are being undertaken to make biocrude produced from *Chlorella* species of microalgae an economically competitive substitute for conventional petroleum. In this work, literature research was conducted to bring together several studies that have already been published. Consequently, all the works done to present that were brought together show that significant areas were not covered in the work the researchers carried out, the data they obtained from their experiment, or in the analysis of their results. Most of the authors reported the effects of operating conditions on the percentage yield of the biocrude from the biomass of *Chlorella* species, the biocrude quality, the elemental percentage composition of the biocrude, and its heating value. However, attention was not focused on how the relevant chemical and biochemical compositions can be modified to improve the biocrude yield and quality while reducing the amount of the compositions that do not favor it. Also, the data currently available for the extraction of biocrude after the hydrothermal liquefaction of *Chlorella* species were obtained by solvent extraction experiments. No comparative study of extraction methods is available, especially mechanical means of extracting the biocrude. The assessments of the composition of the biomass, the operating conditions, and the separation process indicate that hydrothermal liquefaction of *Chlorella* species could become an economically feasible technology for biocrude production, but much work still needs to be done in this area of study for a comprehensive understanding of the conversion process.

REFERENCES

- Aigner, S., Glaser, K., Arc, E., Holzinger, A., Schletter, M., Karsten, U., Kranner, I. (2020). *Adaptation to Aquatic and Terrestrial Environments in Chlorella vulgaris (Chlorophyta)* Frontiers in Microbiology, 585836, 1–14.
- Babich, I.V., Van der Hulst, M., Lefferts, L., Moulijn, J.A., O'Connor, P., Seshan, K. (2011). Catalytic pyrolysis of microalgae to high-quality liquid bio-fuels. *Biomass and bioenergy*, 35, 3199–3207.
- Bach, Q.V., Chen, W.H., Lin, S.C., Sheen, H.K. & Chang, J.S. (2017). Wet torrefaction of microalga *Chlorella vulgaris* ESP-31 with microwave-assisted heating. *Energy Conversion Management*, 141, 163–170.
- Barreiro, D.L., Prins, W., Ronsse, F. & Brilman, W. (2013). Hydrothermal liquefaction (HTL) of microalgae for biofuel production: State of the art review and future prospects'. *Biomass and bioenergy*, 53, 113–127.
- Basar, I.A., Liu, H., Carrere, H., Trably, E. & Eskicioglu, C. (2021). A review on key design and operational parameters to optimize and develop hydrothermal liquefaction of biomass for biorefinery applications. *Green Chemistry*, 23, 1404–1446.
- Biller, P. & Ross, A.B. (2011). Potential yields and properties of oil from the hydrothermal liquefaction of microalgae with different biochemical content. *Bioresource Technology*, 102, 215–225.
- Biller, P., Riley, R. & Ross, A.B. (2011). Catalytic hydrothermal processing of microalgae: decomposition and upgrading of lipids. *Bioresource Technology*, 102, (7), 4841–4848.
- Bito, T., Okumura, E., Fujishima, M., Watanabe, F. (2020). Potential of *Chlorella* as a Dietary Supplement to Promote Human Health. *Nutrients*, 12, (2524), 1–21.
- Bito, T., Tanioka, Y., Watanabe, F. (2018). Characterization of vitamin B₁₂ compounds from marine foods. *Fisheries science*, 84, 747–755.
- Cantero, D.A., Sánchez Tapia, Á., Bermejo, M.D. & Cocero, M.J. (2015). Pressure and temperature effect on cellulose hydrolysis in pressurized water. *Chemical Engineering Journal*, 276, 145–154.
- Cao, L., Zhang, C., Chen, H., Tsang, D.C.W., Luo, G., Zhang, S., & Chen, J. (2017). Hydrothermal liquefaction of agricultural and forestry wastes: state-of-the-art review and future prospects. *Bioresource Technology*, 196, 1–45.
- Chakinala, A.G., Brilman, D.W.F., van Swaaij, W.P.M. & Kersten, S.R.A. (2009). Catalytic and non-catalytic supercritical water gasification of microalgae and glycerol. *Industrial & Engineering Chemistry Research*, 49, 1113–1122.
- Chakraborty, M., McDonald, A.G., Nindo, C. & Chen, S. (2013). An α -glucan isolated as a co-product of biofuel by hydrothermal liquefaction of *Chlorella sorokiniana* biomass. *Algae Research*, 1–7.
- Chang, Y.M., Tsai, W.T. & Li, M.H. (2014). Characterization of activated carbon prepared from *Chlorella*-based algal residue. *Bioresource Technology*, 1–22.
- Changi, S.M., Faeth, J.L., Mo, N. & Savage, P.E. (2015). Hydrothermal Reactions of Biomolecules Relevant for Microalgae Liquefaction'. *Industrial and Engineering Chemistry Research*, 54, (47), 11733–11758.
- Chen, W. Chu, Y. Liu, J. & Chang, J. (2018). Thermal degradation of carbohydrates, proteins and lipids in microalgae analyzed by evolutionary computation. *Energy Conversion and Management*, 160, 209–219.
- Chen, W.H., Lin, B.J., Huang, M.Y., Chang, J.S., (2014). Thermochemical conversion of microalgal biomass into biofuels: A review. *Bioresource Technology*, 184, (910), 314–327.
- Coronado-Reyes, J.A., Salazar-Torres, J.A., Juárez-Campos, B. & González-Hernández, J.C. (2022). *Chlorella vulgaris*, a microalgae important to be used in Biotechnology: a review. *Food Science and Technology*, 42, (37320), 1–11.
- Déniel, M., Haarlemmer, G., Roubaud, A., Weiss-Hortala, E. & Fages, J. (2015). Energy valorisation of food processing residues and model compounds by hydrothermal liquefaction. *Renewable Sustainable Energy Reviews*, 54, 1632–1652.
- Dimitriadis, A. & Bezergianni, S. (2017). Hydrothermal liquefaction of various biomass and waste feedstocks for biocrude production: A state of the art review. *Renewable and Sustainable Energy Reviews*, 68, (1), 113–125.
- Dote, Y., Sawayama, S., Yokoyama, S., Inoue, S., Minowa, T. & Yokoyama, S. (1994). Recovery of liquid fuel from hydrocarbon-rich thermochemical microalgae liquefaction. *Fuel*, 73, 1855–1857.
- Dou, B., Zhang, H., Song, Y., Zhao, L., Jiang, B., He, M., Ruan, C., Chen, H. & Xu, Y. (2019) Hydrogen production from the thermochemical conversion of biomass: issues and challenges. *Sustainable Energy & Fuels*, 3, 314–342.
- Duan, P., Jin, B., Xu, Y., Yang, Y., Bai, X., Wang, F. & Miao, J. (2013). Thermo-chemical conversion of *Chlorella pyrenoidosa* to liquid biofuels. *Bioresource Technology*, 133, 197–205.
- Eboibi, B.E. (2018). Assessing Yield and Properties of Distillate from Biocrude and Blend after Hydrothermal Liquefaction of Microalgae. *Journal of Applied Sciences and Environmental Management*, 22, (6), 949–958.
- Eboibi, B.E. 2019, 'Impact of time on yield and properties of biocrude during downstream processing of product mixture derived from hydrothermal liquefaction of microalga. *Biomass Conversion Biorefinery*, 9, 379–387.
- El-Dalatony, M.M., Salama, E.S., Kurade, M.B., Hassan, S.H.A., Oh, S.E., Kim, S., Jeon, B.H. (2017) Utilization of Microalgal Biofractions for Bioethanol, Higher Alcohols, and Biodiesel Production: A Review. *Energies*, 10, (2110).
- El-Sheekh, M., Abu-Faddan, M., Abo-Shady, A., Nassar, M.Z.A. & Labib, W. (2020). Molecular identification, biomass, and biochemical composition of the marine chlorophyte *Chlorella* sp. MF1 isolated from Suez Bay. *Journal of Genetic Engineering and Biotechnology*, 18, (1), 1–10.
- Embaby, M.A., Haggag, E.A., El-Sheikh, A.S. & Marrez, D.A. (2022). Biosorption of Uranium from aqueous solution by green microalga *Chlorella sorokiniana*. *Environmental Science and Pollution Research*, 29, 58388–58404.
- Fan, S., Zhang, P., Li, F., Jin, S., Wang, S. & Zhou, S. (2016). A Review of Lignocellulose change during Hydrothermal Pretreatment for Bioenergy Production. *Current organic chemistry*, 20, (26), 2799–2809.
- Fan, Y., Hornung, U., Dahmen, N. & Kruse, A. (2018). Hydrothermal liquefaction of protein-containing biomass: study of model compounds for Maillard reactions. *Biomass Conversion Biorefinery*, 8, 909–923.
- Ferreira, A.F. & Soares Dias, A.P. (2020). Pyrolysis of Microalgae Biomass over Carbonate Catalysts. *Journal of Chemical Technology & Biotechnology*, 1–44.
- Figueira, C.E., Moreira, P.F., Giudici, R. (2015). Thermogravimetric analysis of the gasification of microalgae *Chlorella vulgaris*. *Bioresource Technology*, 198, 717–724.
- Gai, C., Zhang, Y., Chen, W.T., Zhang, P., Dong, Y., (2015). An investigation of reaction pathways of hydrothermal liquefaction using *Chlorella pyrenoidosa* and *Spirulina platensis*. *Energy Conversion Management*, 96, 330–339.
- Gao, F., Yang, Z.Y., Zhao, Q.L., Chen, D.Z., Li, C., Liu, M., Yang, J.S., Liu, J.Z., Ge, Y.M. & Chen, J.M. (2021). Mixotrophic cultivation of microalgae coupled with anaerobic hydrolysis for sustainable treatment of municipal wastewater in a hybrid system of anaerobic

- membrane bioreactor and membrane photobioreactor. *Bioresource Technology*, 337, (125457), 1–9.
- Giang, T.T., Lunprom, S., Liao, Q., Reungsang, A. & Salkkam, A. (2019). Improvement of hydrogen production from *Chlorella* sp. biomass by acid-thermal pretreatment. *Biochemistry, Biophysics and Molecular Biology*, 7, (6637).
- Gita, S., Shukla, S.P., Saharan, N., Prakash, C. & Deshmukhe, G. (2019). *Toxic Effects of Selected Textile Dyes on Elemental Composition, Photosynthetic Pigments, Protein Content and Growth of a Freshwater Chlorophycean Alga Chlorella vulgaris*. Bulletin of Environmental Contamination and Toxicology, 1–7.
- Gollakota, A.R.K., Kishore, N., & Gu, S (2018). A review on hydrothermal liquefaction of biomass. *Renewable and Sustainable Energy Reviews*, 5.
- Guo, B., Walter, V., Hornung, U. & Dahmen, N. (2019). Hydrothermal liquefaction of *Chlorella vulgaris* and *Nannochloropsis gaditana* in a continuous stirred tank reactor and hydrotreating of biocrude by nickel catalysts. *Fuel Processing Technology*, 191, 168–180.
- Guo, Y., Yeh, T., Song, S., Xu, D., Wang, S. (2015). A review of bio-oil production from hydrothermal liquefaction of algae. *Renewable and Sustainable Energy Reviews*, 48, 776–790.
- Hao, B., Xu, D., Jiang, G., Sabri, T.A., Jing, Z., Guo, Y. (2021). Chemical reactions in hydrothermal liquefaction of biomass and in catalytic hydrogenation upgrading of biocrude. *Green Chemistry*, 1–30.
- Haque, T.M.A., Perez, M., Brdecka, M., Salas, V.D. & Jang, B. (2022) Effects of Plasma Modification and Atmosphere on the Catalytic Hydrothermal Liquefaction of *Chlorella*. *Industrial & Engineering Chemistry Research*, 1–11.
- He, Z., Xu, D., Liu, L., Wang, Y., Wang, S., Guo, Y. & Jing, (2018). Product characterization of multi-temperature steps of hydrothermal liquefaction of *Chlorella* microalgae. *Algae Research*, 33, 8–15.
- He, Z., Xu, D., Wang, S., Zhang, H. & Jing, Z. (2018). Catalytic upgrading of water-soluble bio-crude from hydrothermal liquefaction of *Chlorella*. *Energy & Fuels*, 1–12.
- Hom-Diaz, A., Llorca, M., Rodriguez-Mozaz, S., Vicent, T., Barcelo, D. & Blaquez P. (2015). Microalgae cultivation on wastewater digestate: β -estradiol and 17 α -ethynylestradiol degradation and transformation products identification. *Journal of Environmental management*, 155, 106–113.
- Hossain, N., Mahlia, T.M.I. & Saidur, R. (2019). Latest development in microalgae-biofuel production with nano-additives. *Biotechnology for Biofuels*, 12, (125).
- Hu, Y., Gong, M., Feng, S., Xu, C.C. & Bassi, A. (2019). A review of recent developments of pre-treatment technologies and hydrothermal liquefaction of microalgae for bio-crude oil production. *Renewable and Sustainable Energy Reviews*, 101, 476–492.
- Huang, Y., Xiong, X., Liao, Q., Fu, Q., Xia, A., Zhu, X. & Sun, Y. (2016). Comparison of *Chlorella vulgaris* biomass productivity cultivated in biofilm and suspension from the aspect of light transmission and microalgae affinity to carbon dioxide. *Bioresources Technology*, 1–32.
- Jabeen, S., Gao, X., Altarawneh, M., Hayashi, J., Zhang, M. & Dlugogorski, B.Z. (2019). Analytical Procedure for Proximate Analysis of Algal Biomass: Case Study for *Spirulina* and *Chlorella*. *Energy & fuels*, 1–29.
- Jaiswal, K.K., Banerjee, I., Singh, D., Sajwan, P. & Chhetri, V. (2020). Ecological stress stimulus to improve microalgae biofuel generation: a review. *Octa Journal of Biosciences*, 8, (1), 48–54.
- Jazrawi, C., Biller, P., He, Y., Montoya, A., Ross, A.B., Maschmeyer, T. & Haynes, B.S. (2015). Two-stage hydrothermal liquefaction of a high-protein microalga. *Algal Research*, 8, 15–22.
- Jena, U., McCurdy, A.T., Warren, A., Summers, H., Ledbetter, R.N. & Hoekman S.K. (2015). Oleaginous yeast platform for producing biofuels via co-solvent hydrothermal liquefaction. *Biotechnology Biofuels*, 8, 167.
- Jin, M., Oh, Y.K., Chang, Y.K. & Choi, M. (2017). Optimum Utilization of Biochemical Components in *Chlorella* sp. KR1 via Subcritical Hydrothermal Liquefaction. *ACS Sustainable Chemistry & Engineering*, 1–9.
- Khoo, C.G., Lamb, M.K., Mohamed, A.R. & Lee, K.T. (2020). Hydrochar production from high-ash low-lipid microalgal biomass via hydrothermal carbonization: Effects of operational parameters and products characterization. *Environmental Research*, 188, (109828), 1–12.
- Kumar, A., Guria, C., & Pathak, A.K. (2018). Optimal cultivation towards enhanced algae biomass and lipid production using *Dunaliella tertiolecta* for biofuel application and potential CO₂ biofixation: Effect of nitrogen deficient fertilizer, light intensity, salinity and carbon supply strategy. *Energy*, 1–51.
- Kumar, G. Shobana, S., Chen, W.H., Bach, Q.V., Kim, S.H., Atabani, A.E. & Chang, J.S. (2016). A review of thermochemical conversion of microalgal biomass for biofuels: Chemistry and processes. *Green Chemistry*, 1–75.
- Lizzul, A., Lekuona-Amundarain, A., Purton, S., Campos, L. (2018). Characterization of *Chlorella sorokiniana*, UTEX 1230', *Biology*, vol. 7, no. 25, pp. 1–12.
- López-González, D., Fernandez-Lopez, M., Valverde, J.L. & Sanchez-Silva, L. (2014). Pyrolysis of three different types of microalgae: kinetic and evolved gas analysis. *Energy*, 73, 33–43.
- Mathimani, T. & Mallick, N. (2019). A review on the hydrothermal processing of microalgal biomass to bio-oil - Knowledge gaps and recent advances. *Journal of Cleaner Production*, 217, 69–84.
- Miao, C., Chakraborty, M. & Chen, S. (2012). Impact of reaction conditions on the simultaneous production of polysaccharides and bio-oil from heterotrophically grown *Chlorella sorokiniana* by a unique sequential hydrothermal liquefaction process. *Bioresource Technology*, 110, 617–627.
- Mobin S.M.A., Chowdhury, H. & Alam, F. (2019). *Commercially important bioproducts microalgae and their current applications – A review*. Energy Procedia, 2nd International conference on energy and power, Sydney, Australia, pp 1–9.
- Niccolai, A., Zittelli, G.C., Rodolfi, L., Biondi, N. & Tredici, M.R. (2019). Microalgae of interest as food source: Biochemical composition and digestibility. *Algae Research*, 42.
- Nwanya, K.O., Okoye, A.C. & Ajiwe, I.E. (2021). Biodiesel Potentials of *Chlorella vulgaris* Oil. *Nigerian Research Journal of Chemical Sciences*, 9, (2), 36–46.
- Peng, G., Vogel, F., Refardt, D., Ludwig, C. (2017) Catalytic Supercritical Water Gasification: Continuous Methanization of *Chlorella vulgaris*. *Industrial Engineering Chem. Res.* 56, 6256–6265.
- Peng, H., Wei, D., Chen, G. & Chen, F. (2016). Transcriptome analysis reveals global regulation in response to CO₂ supplementation in oleaginous microalga *Coccomyxa subellipsoidea* C-169. *Biotechnology Biofuels*, 9, (151).
- Phusunti, N., Phetwarotai, W., Tekasakul, S. (2017). Effects of torrefaction on physical properties, chemical composition and reactivity of microalgae. *Korean Journal of Chemical Engineering*, 1–8.
- Piasecka, A., Baier, A. (2022). Metabolic and Proteomic Analysis of *Chlorella sorokiniana*, *Chloroidium saccharophilum*, and *Chlorella vulgaris*

- Cells Cultured in Autotrophic, Photoheterotrophic, and Mixotrophic Cultivation Modes. *Molecules*, 27 (4817), 2–16.
- Qiu, Y., Aierzhati, A., Cheng, J., Guo, H., Yang, W. & Zhang, Y. (2019). Biocrude Oil Production through the Maillard Reaction between Leucine and Glucose during Hydrothermal Liquefaction. *Energy & Fuels*, 33, (9), 8758–8765.
- Rabaçal, M., Ferreira, A.F., Silva, C.A.M., Costa, M. (2017). *Lecture Notes in Energy, Biomass Availability, Potential and Characteristics', Biorefineries*, 57, Chapter 2, 21–54.
- Raheem, A., Azlina, W.A.K.G., Taufiq Yap, Y.H., Danquah, M.K., Harun, R. (2015). Thermochemical conversion of microalgal biomass for biofuel production. *Renewable Sustainable Energy Review*, 49, 990–999.
- Rani, K., Sandal, N. & Sahoo, P.K. (2018). A comprehensive review on chlorella- its composition, health benefits, market and regulatory scenario. *The Pharma Innovation Journal*, 7, (7), 584–589.
- Rani, S. & Ojha, C.S.P. (2021). Chlorella sorokiniana for integrated wastewater treatment, biomass accumulation and value-added product estimation under varying photoperiod regimes: A comparative study. *Journal of Water Process Engineering*, 39, (101889), 1–9.
- Rinanti, A. & Purwadi, R. (2019). Increasing carbohydrate and lipid productivity in tropical microalgae biomass as a sustainable biofuel feed stock. *Energy Procedia*, 158, 1215–1222.
- Rinanti, A. & Purwadi, R. (2019). The potential of tropical microalgae as flocculant in harvesting process. *International Journal of GEOMATE*, 16, (56), 165–170.
- Rizzo, A.M., Prussi, M., Bettucci, L., Libelli, I.M., Chiamonti, D. (2013). Characterization of microalga Chlorella as a fuel and its thermogravimetric behavior. *Applied Energy*, 102, 24–31.
- Ross, A.B., Biller, P., Kubacki, M.L., Li, H., Lea-Langton, A. & Jones, J.M. (2010). Hydrothermal processing of microalgae using alkali and organic acids. *Fuels*, 89, (9), 2234–2243.
- Ru, J., Huo, Y. & Yang, Y. (2020). Microbial Degradation and Valorization of Plastic Wastes. *Frontiers in Microbiology*, 11, (442), 1–20.
- Safar, H., Nørregaard, P.U., Ljubic, A., Møller, P., Holdt, S.L. & Jacobsen, C. (2016). Enhancement of Protein and Pigment Content in Two Chlorella Species Cultivated on Industrial Process Water. *Journal of Marine Science and Engineering*, 4, (84), 1–15.
- Sajjadi, B., Chen, W.-Y., Raman, A.A.A., Ibrahim, S. (2018). Microalgal lipid and biomass for biofuel production: A comprehensive review on lipid enhancement strategies and their effects on fatty acid composition. *Renewable and Sustainable Energy Reviews*, 97, 200–232.
- Sakarika, M. & Kornaros, M. (2017). Kinetics of growth and lipids accumulation in Chlorella vulgaris during batch heterotrophic cultivation: Effect of different nutrient limitation strategies. *Bioresource Technology*, 243, 356–365.
- Shanmugam, S., Mathimani, T., Anto, S., Sudhakar, M.P., Kumar, S.S., Pugazhendhi, A. (2020). Cell density, Lipidomic profile, and fatty acid characterization as selection criteria in bioprospecting of microalgae and cyanobacterium for biodiesel production. *Bioresource Technology*, 304, (123061).
- Song, W., Wang, S., Guo, Y. (2015). A Review of the Biofuel yield in hydrothermal liquefaction of different microalgae. *International Conference on Advances in Energy and Environmental Science*, 1504–1509.
- Tang, G., Suter, P.M. (2011). Vitamin A, Nutrition, and Health Values of Algae: Spirulina, Chlorella, and Dunaliella. *Journal of Pharmacy and Nutrition Sciences*, 1, 111–118.
- Tarek, D., Ahmed, M.M., Hussein, H.S., Zeyad, A.M., Al-Enizi, A.M., Yousef, A., Ragab, A. (2022). Building envelope optimization using geopolymers to improve the energy efficiency of residential buildings in hot arid regions. *Case Studies in Construction Materials*, 17, 01657.
- Terrell, E., Dellon, L.D., Dufour, A., Bartolomei, E., Broadbelt, L.J. & Garcia-Perez, M. (2020). A Review on Lignin Liquefaction: Advanced Characterization of Structure and Microkinetic Modeling. *Industrial and engineering chemistry research*, 59, (30), 56–555.
- Tian, C., Li, B., Liu, Z., Zhang, Y. & Lu, H. (2014). Hydrothermal liquefaction for algal biorefinery: A critical review. *Renewable and Sustainable Energy Reviews*, 38, 933–950.
- Tian, S.L., Khan, A., Zheng, W.N., Song, L., Liu, J.H., Wang, X.Q., Li, L. (2022). Effects of Chlorella extracts on growth of Capsicum annuum L. seedlings. *Scientific report*, 12, 15455.
- Wang, J., Xiao, B., Liu, S., Hu, Z., He, P., Guo, D., Hu, M., Qi, F., Luo, S., (2013). Catalytic steam gasification of pig compost for hydrogen-rich gas production in a fixed bed reactor. *Bioresource Technology*, 133, 127–133.
- Wang, X., Lin, L., Lu, H., Liu, Z., Duan, N., Dong, T., Xiao, H., Li, B. & Xu, P. (2018). Microalgae cultivation and culture medium recycling by a two-stage cultivation system. *Frontiers of Environmental Science & Engineering*, 12, (14).
- Watson, J., Wang, T., Si, B., Chen, W.T., Aierzhati, A. & Zhang, Y. (2020). Valorization of hydrothermal liquefaction aqueous phase: pathways towards commercial viability. *Progress in Energy and Combustion Science*, 17, (100819).
- Xu, Y., Hu, Y., Peng, Y., Yao, L., Dong, Y., Yang, B., Song, R. (2019). Catalytic pyrolysis and liquefaction behavior of microalgae for bio-oil production. *Bioresource Technology*, 1–37.
- Xue, Y., Chen, H., Zhao, W., Yang, C., Ma, P. & Han, S. (2016). A review on the operating conditions of producing biooil from hydrothermal liquefaction of biomass. *International Journal of Energy Research*, 40, 865–877.
- Yang, J.H., Shin, H.S., Ryu, Y.J. & Lee, C.G. (2018). Hydrothermal liquefaction of Chlorella vulgaris: Effect of reaction temperature and time on energy recovery and nutrient recovery. *Journal of Industrial and Engineering Chemistry*, pp. 1–7.
- Yu, G., Zhang, Y., Schideman, L., Funk, T.L. & Wang, Z. (2011). Hydrothermal liquefaction of low lipid content microalgae into bio-crude oil. *Transaction of the American Society of Agricultural and Biological Engineers*, 54, 239–46.
- Zhang, B., Lin, Q., Zhang, Q., Wu, K., Pu, W., Yang, M. & Wu, Y. (2017). Catalytic hydrothermal liquefaction of Euglena specie microalgae over zeolite catalysts for the production of bio-oil. *Royal society of chemistry advances*, 7, 8944–8951.
- Zhang, C., Wang, C., Cao, G., Chen, W.-H. & Ho, S.H. (2019). Comparison and characterization of property variation of microalgal biomass with non-oxidative and oxidative torrefaction. *Fuel*, 246, 375–385.
- Zhang, J., Chen, W.T., Zhang, P., Luo, Z. & Zhang, Y. (2013). Hydrothermal liquefaction of Chlorella pyrenoidosa in sub- and supercritical ethanol with heterogeneous catalysts. *Bioresources Technology*, 133, 389–397.
- Zhang, J., Luo, Z. & Zhang, Y. (2013). Hydrothermal liquefaction of chlorella pyrenoidosa in water and ethanol. *American Society of Agricultural and Biological Engineers*, 56, (1), 253–259.
- Zhang, J., Zhang, Y. & Luo, Z. (2014). Hydrothermal Liquefaction of Chlorella pyrenoidosa in Ethanol-water for Bio-crude Production. *The 6th International Conference on Applied Energy*, Energy Procedia, 61, 1961–1964.
- Zhang, W., Zhang, P., Sun, H., Chen, M., Lu, S. & Li, P. (2014). Effects of various organic carbon sources on the growth and biochemical composition of Chlorella pyrenoidosa. *Bioresource Technology*, 173, 52–58.
- Zhang, W., Zhang, X., Lei, F., Jiang, J. (2020). Co-production bioethanol and xylooligosaccharides from sugarcane bagasse via autohydrolysis pretreatment. *Renewable Energy*, 1–41.

- Zhang, Y. & Chen, W.T. (2018).Hydrothermal liquefaction of protein-containing feedstocks.*Direct Thermochemical Liquefaction for Energy Applications*, 127–168.
- Zheng, H., Wu, X., Zou, G., Zhou, T., Liu, Y. &Ruan, R. (2019). Cultivation of Chlorella vulgaris in manure-free piggery wastewater with high-strength ammonium for nutrients removal and biomass production: Effect of ammonium concentration, carbon/nitrogen ratio and pH.*Bioresource Technology*, 273, 203–211.
- Zhu, X., Phinney, D.M., Paluri, S.&Heldman, D.R.(2018).Prediction of Liquid Specific Heat Capacity of Food Lipids.*Journal of food sciences*, 0, (0), 1–6.

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